

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION  
International Bureau



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification 6 :</b> <b>C12N 15/87, A61K 48/00, 47/48, C12Q 1/70 // C12N 15/86</b>	<b>A1</b>	<b>(11) International Publication Number:</b> <b>WO 97/05266</b> <b>(43) International Publication Date:</b> 13 February 1997 (13.02.97)
<b>(21) International Application Number:</b> PCT/NL96/00302 <b>(22) International Filing Date:</b> 25 July 1996 (25.07.96) <b>(30) Priority Data:</b> 95202040.2 25 July 1995 (25.07.95) EP <b>(34) Countries for which the regional or international application was filed:</b> NL et al. <b>(71) Applicant (for all designated States except US):</b> INTROGENE B.V. [NL/NL]; Lange Kleiweg 151, NL-2288 GJ Rijswijk (NL). <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only):</b> VALERIO, Domenico [NL/NL]; Lange Kleiweg 151, NL-2288 GJ Rijswijk (NL). VAN BEUSECHEM, Victor, Willem [NL/NL]; Lange Kleiweg 151, NL-2288 GJ Rijswijk (NL). <b>(74) Agent:</b> SMULDERS, Th., A., H., J.; Vereenigde Octrooibureaux, Nieuwe Parklaan 97, NL-2587 BN The Hague (NL).		<b>(81) Designated States:</b> AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>With international search report.</i>
<b>(54) Title:</b> METHODS AND MEANS FOR TARGETED GENE DELIVERY  <b>(57) Abstract</b>  The present invention provides a method of producing viral gene delivery vehicles which can be transferred to preselected cell types by using targeting conjugates. The gene delivery vehicles comprise: 1) the gene of interest (to be delivered); 2) a viral capsid or envelope carrying a member of a specific binding pair, the counterpart of which is not directly associated with the surface of the target cell. These vehicles can be made unable to bind to their natural receptor on the cell. The targeting conjugates are composed of the counterpart member of the specific binding pair linked to a targeting moiety which is a cell-type specific ligand (or fragments thereof). The number of the specific binding pair present on the viral vehicles can be for example an immunoglobulin binding moiety (e.g. capable of binding to a Fc fragment, protein A, protein G, FcR or an anti-Ig antibody), or biotin, avidin or streptavidin. The outer membrane or capsid of the virus may contain a substance which mediates entrance of the gene delivery vehicle into the target cell. Due to the specificity of the ligand, the high affinity of the binding pair and to the inability of the gene delivery vehicle to be targeted when used alone, the universality of the method for gene delivery, together with its high cell type selectivity can easily be achieved by the use of various adequate targeting conjugates.		

**FOR THE PURPOSES OF INFORMATION ONLY**

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AM	Armenia	GB	United Kingdom	MW	Malawi
AT	Austria	GE	Georgia	MX	Mexico
AU	Australia	GN	Guinea	NE	Niger
BB	Barbados	GR	Greece	NL	Netherlands
BE	Belgium	HU	Hungary	NO	Norway
BF	Burkina Faso	IE	Ireland	NZ	New Zealand
BG	Bulgaria	IT	Italy	PL	Poland
BJ	Benin	JP	Japan	PT	Portugal
BR	Brazil	KE	Kenya	RO	Romania
BY	Belarus	KG	Kyrgyzstan	RU	Russian Federation
CA	Canada	KP	Democratic People's Republic of Korea	SD	Sudan
CF	Central African Republic	KR	Republic of Korea	SE	Sweden
CG	Congo	KZ	Kazakhstan	SG	Singapore
CH	Switzerland	LI	Liechtenstein	SI	Slovenia
CI	Côte d'Ivoire	LK	Sri Lanka	SK	Slovakia
CM	Cameroon	LR	Liberia	SN	Senegal
CN	China	LT	Lithuania	SZ	Swaziland
CS	Czechoslovakia	LU	Luxembourg	TD	Chad
CZ	Czech Republic	LV	Latvia	TG	Togo
DE	Germany	MC	Monaco	TJ	Tajikistan
DK	Denmark	MD	Republic of Moldova	TT	Trinidad and Tobago
EE	Estonia	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	UG	Uganda
FI	Finland	MN	Mongolia	US	United States of America
FR	France	MR	Mauritania	UZ	Uzbekistan
GA	Gabon			VN	Viet Nam

Title: Methods and means for targeted gene delivery.

Field of the Invention:

The present invention relates to the field of providing cells with additional genetic information through recombinant DNA techniques. It especially relates to methods and means of providing specific groups of cells with additional genetic information, in particular in the context of gene therapy.

Gene therapy is a recently developed concept which has a very broad range of applications, ranging from supplementing deficiencies in a mammals' set of proteins, usually resulting from genetic disorders, to the treatment of cancer, (auto-)immune diseases or (viral) infections, usually by eliminating or suppressing the responsible set of cells or organisms.

One of the main problems in gene therapy is delivering the genetic material to the target cells and not to other cells. Another problem in gene therapy is that certain cell types are extremely refractory to current gene transfer techniques.

The present invention relates generally to gene delivery vehicles and their use in gene therapy, and more particularly, to recombinant viruses which can be targeted to susceptible target cells.

Retroviruses are RNA viruses which efficiently integrate their genetic information into the genomic DNA of infected cells via a reverse-transcribed DNA intermediate. This property of their life-cycle and the fact that parts of their genetic material can be replaced by foreign DNA sequences make retroviruses one of the most promising vectors for the delivery of genes in human gene therapy procedures, most notably for gene therapies which rely on gene transfer into dividing tissues. Most retroviral vector systems are based on mouse retroviruses and consist of two components, i.e. (i) the recombinant retroviral vector

carrying the foreign sequences of interest, and (ii) so-called packaging cells expressing the structural viral proteins of which the encoding sequences are lacking in the retroviral vector. Expression of (i) in (ii) results in the production of recombinant retroviral particles capable of transducing susceptible target cells.

The infectivity and host cell range of the retrovirus particle is conferred by an envelope glycoprotein which specifically binds to a receptor molecule on the target cell membrane. The envelope glycoprotein of all known retroviruses consists of two associated peptides, which are derived by proteolytic cleavage from the same precursor protein encoded by the retroviral *env* gene (Dickson et al. in Weiss et al. (ed.) Molecular biology of tumor viruses (1984), Cold Spring Harbor Press, pp.513-648). The amino terminal peptide encompasses the specific binding site for its receptor on the target cell membrane, thus determining the virus host range (Hunter and Swanstrom, Curr. Top. Microbiol. Immunol. 157(1990):187). The carboxy terminal peptide, which contains trans-membrane anchor sequences, is assumed to account for the selective uptake of the envelope glycoprotein in the virus particle and to mediate fusion between the virus membrane and - depending on the type of virus - the plasma membrane or intracellular vesicle membrane of the target cell.

Several envelope glycoprotein variants with different infection spectra for mammalian cells have been identified. All known *env* variants have a rather broad infection spectrum in common. Here lies one of the major shortcomings of current recombinant retrovirus technology. In numerous gene therapy applications targeted delivery of genes into defined cells would be desired, most notably in the case of *in vitro* gene transfer into cell types present with low abundance in cell mixtures and in approaches for *in vivo* gene transfer into cells in a living mammalian body. Conventional gene transfer techniques have the disadvantage in such applications of low efficiency of gene transfer to

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

desired target cells, because the gene transfer vehicles are taken up by other cells as well. In addition, the cell mixture or living mammalian body may contain cells to which gene transfer is absolutely undesired. E.g., genes providing protection against chemotherapeutic drugs should not be transferred into malignant cells. Therefore, increasing attention is focused on devising procedures to limit the retrovirus infection spectrum. By employing particular env variants the transduction spectrum can be limited to some extent, but true specificity for most target cells of interest can not be obtained this way. On the other hand, on the surface of some specific cell types the expression of receptors for retroviral entry is extremely low. An important example of a cell type with low retrovirus receptor expression is the pluripotent stem cell of the hematopoietic system (Orlic et al., Blood 86 suppl 1 (1995):628a). A preferred strategy to accomplish targeted delivery is to direct the retrovirus particle to cell membrane molecules differing from the natural envelope glycoprotein receptor, said molecule being specifically expressed on the membrane of the desired target cell. Present ideas about how this could be done include:

1. direct chemical coupling of a ligand for a target cell molecule to the viral envelope glycoprotein (Neda et al, J. Biol. Chem. 266(1991):14143),
2. bridging the viral envelope glycoprotein to a molecule on the target cell through a complex of two antibodies, one directed against the viral envelope glycoprotein and the other against the molecule on the target cell (Goud et al., Virology 163(1988):251; Roux et al., Proc. Natl. Acad. Sci. USA 86(1989):9079; Etienne-Julan et al., J. Gen. Virol. 73(1992):3251),
3. bridging the viral envelope glycoprotein to a molecule on the target cell through a complex of an antibody directed against the viral envelope glycoprotein and a peptide ligand for the molecule on the target cell (Etienne-Julan et al., J. Gen. Virol. 73(1992):3251)

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

4. replacing the specific binding site of the viral envelope glycoprotein for its receptor by a peptide ligand for a target cell surface molecule,
5. co-expression on the virus membrane of the natural viral envelope glycoprotein and a ligand for a target cell surface molecule (Young et al., Science 250(1990):1421), and
6. co-expression on the virus membrane of the natural viral envelope glycoprotein and an altered viral envelope glycoprotein in which the specific binding site for its receptor has been replaced by a peptide ligand for a target cell surface molecule (Young et al., Science 250(1990):1421; Russell et al., Nucl. Acid Res. 5(1993):1081; Chu et al., Gene Ther. 1(1994):292; Kasahara et al., Science 266(1994):1373; Chu and Dornburg, J. Virol. 69(1995):2659).

Monoclonal antibodies or fragments thereof, exhibiting high specificity and affinity for the target cell specific molecule, are amongst the preferred ligands for targeted delivery. Approaches nos. 2 and 3 mentioned above rely on antibodies and promising tools for use in the approaches nos. 4 and 6 are chimaeric molecules between viral envelope glycoproteins and single-chain antibody fragments of the variable antigen-binding domain of immunoglobulins (scFv) (Russell et al., Nucl. Acid Res. 5(1993):1081; Chu et al., Gene Ther. 1(1994):292; Chu and Dornburg, J. Virol. 69(1995):2659). A chimaeric molecule of such an scFv fragment and a different membrane anchoring protein than the viral envelope glycoprotein could be used for approach no. 5.

An important limitation of all these previously invented approaches is that a new virus with a specific targeting ligand (chemically or genetically modified envelope glycoprotein, or co-expressed ligand) or a new specific dual-antibody complex has to be made for each

target cell type. The present invention bypasses this limitation.

Adenovirus capsids are regular icosahedrons composed of 252 subunits, of which 240 are so-called hexons and 12 are so-called pentons. The pentons are located at the vertices of the icosahedron. They contain a penton base on the surface of the capsid which is composed of five molecules of a 85 kD polypeptide. A fiber composed of a homotrimer of 62 kD polypeptides projects from the penton base outward. The fiber protein is responsible for attachment of the adenovirus to its receptor (reviewed by Horwitz, in: Virology, 2nd edition (Fields et al, ed), Raven Press, New York, 1990, pp.1679-1721). By exchanging fiber protein domains from two adenoviruses of different serotype, Stevenson et al. (J. Virol. 69(1995):2850) have shown that the receptor specificity is determined by the head domain of the fiber protein.

The adeno-associated virus (AAV) capsid is comprised of three viral proteins (VP): VP-1, VP-2, and VP-3. These proteins have molecular masses of 87 kD, 73 kD, and 62 kD, respectively. In mature virions VP-1, VP-2 and VP-3 are found at relative abundance of approximately 1 : 1 : 10. In vitro, the three proteins assemble spontaneously into virion-like structures. It appears, therefore, that capsid formation in infected cells proceeds independent of viral DNA synthesis (reviewed by Kotin, Hum. Gene Ther. 5(1994):793). It has been shown possible to insert sequences into the genes encoding the capsid proteins, resulting in the exposure of His-residues on the surface of intact AAV capsids. Consequently, these altered AAV virions were able to bind to a nickel-column (unpublished results from the group of Dr. R. Samulski, Univ. of North Carolina, Chapel Hill, NC). The present invention makes use of the possibility to insert sequences in said capsid genes.

The present invention thus provides a gene delivery vehicle for delivering genetic material to a target cell, comprising an expressible recombinant nucleic acid molecule

encoding a gene product of interest, a capsid or an envelope derived from a virus, and a member of a specific binding pair, the counterpart of which is not directly associated with the surface of the target cell.

5        This gene delivery vehicle itself is preferably incapable of specifically binding to a target cell, meaning that it is no longer an infectious virus particle. Instead of having its regular infectivity it is provided with a member of a specific binding pair, either as a part of its  
10 envelope or as a part of its capsid. This gene delivery vehicle in itself cannot be targeted to a target cell, but it is a novel and inventive intermediate in the process of preparing a targeting gene delivery vehicle, which together with a targeting component is capable of delivering a gene  
15 to target cells only, or at least of delivering genes to the target cells in a far greater amount than to non-target cells (or other targets, such as infectious organisms), or of delivering a gene to cells previously difficult to transduce as a result of low viral receptor expression. The  
20 invention thus also provides a targeting component which is a conjugate for use in targeting a gene delivery vehicle as disclosed above to a target cell, which conjugate comprises the counterpart of the member of a specific binding pair (which member is present on or in the gene delivery vehicle,  
25 which counterpart is coupled to a targeting moiety being capable of binding to a target molecule associated with the surface of a target cell.

Thus the conjugate has dual specificity. On the one side the counterpart recognizes the member of a specific  
30 binding pair present on or in the gene delivery vehicle and on the other side it recognizes a target molecule associated with the surface of the target cell. When such a conjugate is coupled to a gene delivery vehicle a targeting gene delivery vehicle is obtained. Thus the invention also  
35 provides a targeting gene delivery vehicle, comprising a gene delivery vehicle according as disclosed hereinabove, coupled to a conjugate according also described above.



The main advantage of the gene delivery vehicles according to the invention is that they have a recognizable moiety, which is independent from the target cell the genes have to be delivered to. This means that they can be  
5 targeted to many different target cells, by using different conjugates which all recognize the vehicle (a constant part of the conjugate) and which recognize a number of different targets associated with target cells on the other side. Thus the gene of interest can be delivered to many different  
10 cells, without having to prepare a new vehicle.

A similar approach has been disclosed and is described above as approach no.3. In this approach however, the vehicle has not been provided with an additional member of a specific binding pair, but a viral antigen (the envelope  
15 glycoprotein) itself is the member of a specific binding pair being recognized by an antibody. Apart from the drawback of having to make a new specific dual-antibody complex for every delivery system, an even more important drawback is that all antigens have to be bound to an  
20 antibody, because otherwise the vehicle will retain its capability of infecting its regular host cells, whereas for the gene delivery vehicles according to the invention these glycoproteins are preferably (if not necessarily) not present or altered to impair their normal function.

25 In the following it will become clear that preferred embodiments of the present invention are so called kits (kits of parts) which provide a gene delivery vehicle as one part and at least one targeting moiety as another part.

Another part of the invention therefor is a kit of  
30 parts for delivering genetic material to a target cell, comprising a gene delivery vehicle according to the invention and at least one conjugate of the counterpart member of the specific binding pair coupled to a targeting moiety, said targeting moiety being capable of binding to a  
35 target molecule associated with the surface of a target cell.

The embodiments given above are the general formats of the invention. Thus it is clear that as well a gene delivery vehicle itself, as well as a conjugate, the delivery vehicle coupled to a conjugate and the kit in which both conjugate and delivery vehicle all can be adjusted to the specific embodiments which are recited for the kits only. Thus the specific conjugates without the delivery vehicle and the specific vehicles without the conjugates as well as the specific targeting vehicles are part of this invention as well. For example where a kit of parts is disclosed comprising as a member an immunoglobulin binding protein and as its counterpart an immunoglobulin, this means that the conjugate comprising the immunoglobulin is part of the invention as is the gene delivery vehicle comprising the immunoglobulin binding protein.

The definitions in the claims are themselves clearly understood in the art.

Specific binding pairs for instance are considered to be any two molecules which have a high affinity specific interaction between the two members (herein often referred to as the member and its counterpart). These may be antibody (fragments or derivatives) and the corresponding antigen, receptors and their ligands (in this case especially proteinaceous ligands), well known binding pairs such as avidin/biotin or streptavidin/biotin, peptide structures that can specifically interact in solution and the like. In many instances members of these binding pairs will not be normally present in the envelope or the capsid of a virus and will thus also not be normally present on the gene delivery vehicles according to the invention, For that purpose they may have to be modified in a suitable manner, for instance chemically, or they may need to be present as fusion molecules or hybrid molecules linked to a component which ensures their presence in or on the capsid or envelope. As long as they still have the same function (in kind not in amount) as the original member of the binding pair (i.e. they still bind the counterpart) they should be

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

considered as being a derivative or equivalent of the original member of the binding pair and are thus part of the present invention. The same goes for the counterpart of the member of the specific binding pair and also for the targeting moiety, which is the other side of the conjugate.

Hereinunder a number of possibilities of providing a capsid or an envelope with the function of a member of a specific binding pair are given, exemplified for the preferred vehicles, but skilled artisans will be able to transfer these teachings to other vehicles and/or other members of specific binding pairs as well as other methods of providing these members on envelopes and/or capsids without departing from the present invention.

A conjugate is defined for the purposes of the present invention as Any molecule which has at least two different specific recognition sites, which are somehow linked. This means that it does not necessarily include chemical coupling or fusion proteins, although these are conjugates according to the invention, but that it also includes molecules which normally have this double specific recognition, such as for instance antibodies which recognize an antigen specifically and of which the constant region is specifically recognized by for instance Fc receptors. A nucleic acid encoding a gene of interest should be interpreted as not only encoding proteins, but also antisense nucleic acids and other useful nucleic acids, although these are not usually considered to be genes. When the nucleic acids do encode proteins there are many known genes of interest in the field, particularly in the field of gene therapy. All these genes can be delivered to target cells using the methods and means of the present invention. Of particular interest are suicide genes, such as Herpes Simplex Virus Thymidine Kinase and others disclosed in WO 93/07281.

The target cells to which the genes of interest can be delivered are basically any target cells that have a target molecule associated with their surface with which it is possible to distinguish them from other cells using a

targeting moiety. The ability to distinguish may lie in the abundance of a certain target molecule on a certain subset of cells. Furthermore, it is not necessary for the application of the invention that the nature of the target molecule is known. Targeting moieties can be selected from combinatorial peptide libraries on the basis of differential binding to molecules expressed on the surface of different cell types. Useful combinatorial peptide libraries for the invention include those in which a large variety of peptides is displayed on the surface of filamentous bacteriophages (e.g. Scott and Smith, Science 249(1990):386). Screening for individual library members that interact with desired target cells allows the isolation of the nucleotide sequences encoding suitable peptide structures to be used as a targeting moiety in the invention. For this purpose, libraries displaying scFv variants are particularly useful. Moreover, methods that increase the combinatorial diversity of said libraries make the number of targeting moieties that can be generated for application in the invention almost limitless. Said methods include PCR-based random mutagenesis techniques (Stemmer, Proc. Natl. Acad. Sci. USA 91(1994):10747; Cramer and Stemmer, Biotechniques 18(1995):194). Thus it is understood from the above that the invention discloses methods for specific gene delivery into any target cell that can be phenotypically distinguished from other cells, also when the nature of this distinction has not been revealed. Important target cells are tumor cells, cells of the hematopoietic system, hepatocytes, endothelial cells, lung cells, cells of the central nervous system, muscle cells and cells of the gastro-intestinal tract. Usual target molecules are receptors, surface antigens and the like.

As indicated above the invention also includes a kit of parts wherein the member of a specific binding pair is an immunoglobulin binding moiety and its counterpart is an immunoglobulin or a derivative or a fragment thereof.

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

Preferably the immunoglobulin binding moiety is capable of binding to a constant region of an immunoglobulin. A much preferred immunoglobulin binding moiety is protein A, protein G, or an Fc receptor. One of the most commonly used immunoglobulin-binding proteins is *Staphylococcus aureus* protein A. This 42 kD polypeptide exhibits strong binding to the Fc region of many IgG molecules, including human IgG1, IgG2 and IgG4 and mouse IgG2a and IgG2b, without interfering with the antigen binding site (Surolia et al., Trends Biochem. Sci. 7(1981):74; Lindmark et al, J. Immunol. Meth. 62(1983):1). A wider range of mammalian immunoglobulins is bound by protein G, which is isolated from a group C *Streptococcus* species. It has strong binding capacity for all human and mouse IgG subclasses (Bjork and Kronvall, J. Immunol. 133(1984):969; Sjöbring et al., J. Biol. Chem. 266(1991):399). Protein A and G do not intrinsically bind to mammalian cell surfaces or virus membranes or capsids. Expression of (parts of) these molecules on the surface of gene delivery vehicles will therefore require the generation of hybrid molecules. A hybrid between streptavidin and two of the IgG binding domains of protein A has been made and bound one IgG molecule per hybrid (Sano and Cantor, Bio/Technol. 9(1991):1378).

The mammalian receptors for the Fc domain of immunoglobulins (FcR), however, are trans-membrane molecules (reviewed by Van de Winkel and Capel, Immunol. Today 14(1993):215). These receptors provide a feedback between the humoral and cellular immune responses. Their interaction with immunoglobulins triggers immune functions such as phagocytosis, cytotoxicity, cytokine release and enhancement of antigen presentation. FcR are members of the immunoglobulin superfamily and in humans three classes of receptors for the Fc domain of IgG (FcγR) are recognized (hFcγRI, hFcγRII, and hFcγRIII). hFcγRI is unique in its capacity to bind with high affinity to monomeric IgG ( $K_a = 10^8$ - $10^9$  M<sup>-1</sup>). Its binding is strong to human IgG3, IgG1, and IgG4 (with decreasing affinity), and to mouse IgG2a and

IgG3, whereas human IgG2 and mouse IgG1 and IgG2b are bound much weaker. hFcγRI is constitutively expressed on monocytes and macrophages and its expression can be induced on neutrophils and eosinophils. Three highly homologous genes  
5 for hFcγRI have been identified. Of these, hFcγRIA encodes a trans-membrane molecule, hFcγRIa (α-chain), consisting of three extracellular immunoglobulin-like domains (one of which is not found among other members of the hFcγR-family), a 21 amino acid trans-membrane region, and a charged  
10 cytoplasmic tail of 61 amino acids. On the membrane of some cell types the hFcγRIa α-chain is found associated with a disulfide-linked homodimer of γ-chains, which are also components of other members of the FcR family (Scholl and Geha, Proc. Natl. Acad. Sci. USA 90(1993):8847). There  
15 exists some controversy whether the presence of γ-chains is absolutely essential for membrane expression and ligand binding of the hFcγRIa α-chain (Takai et al, Cell 76(1994):519). The signal transduction after immunoglobulin binding to the α-chain into the interior of the cell is  
20 mediated by the γ-chains; the cytoplasmic domain of the α-chain does not seem to play a role in signal transduction (Indik et al, Exp. Hematol. 22(1994):599).

Because the constant regions of immunoglobulins differ between species, anti-immunoglobulin antibodies can be  
25 generated by cross-species immunization. For several species antibodies have been raised which show specificity to individual immunoglobulin chains or even subclasses. Molecular cloning of the sequences encoding these antibodies offers the possibility to construct scFv, which comprise of  
30 the variable regions of the heavy and light chains of the immunoglobulin molecule linked by a flexible peptide bridge. Such scFv have been shown to be as potent in antigen binding as are immunoglobulin Fab fragments (Bird et al, Science 242(1988):423). Hybrid molecules between scFv and virus  
35 envelope molecules have been made and expressed on the surface of mammalian cells and were shown to be capable of antigen binding and targeted gene delivery (Chu et al, Gene

Ther. 1(1994):292; Chu and Dornburg, J. Virol.  
69(1995):2659).

When the gene delivery vehicle is provided with an immunoglobulin binding moiety, only a suitable antibody (or  
5 a derivative or a fragment (retaining the binding site for the immunoglobulin binding protein)) is needed as the entire conjugate. Thus, when an antibody is available which can distinguish a target cell from other cells, the gene  
10 delivery vehicle is ready to be used to target the gene of interest to said cell using said antibody. Thus for any cell which can be distinguished from other cells using antibodies, a targeted gene delivery system is provided.

The same is true for another embodiment of the present invention, in which the member of a specific binding pair is  
15 biotin and its counterpart is avidin or streptavidin, or in which the member of a specific binding pair is avidin or streptavidin and its counterpart is biotin. In this embodiment the delivery vehicle is provided with the one member of the (strept)avidin/biotin couple and thus the  
20 conjugate comprises the other member.

Thus any targeting moiety that can be biotinylated or can be coupled to (strept)avidin is now usable with the gene delivery vehicle according to the present invention. This makes the number of targeting moieties that can be applied  
25 almost limitless. Any molecule associated with the surface of a target cell for which a specific binding partner is known or can be produced is in principle useful as a target for the presently invented gene delivery system, if in presence or abundance it differs from one subset of cells to  
30 another. Antibodies of course are a good example of suitable targeting moieties of this last embodiment, as they were for the embodiment before that. Thus a preferred embodiment of the present invention encompasses a kit of parts wherein the targeting moiety is an antibody or a fragment or a  
35 derivative thereof, recognizing the target molecule associated with the surface of the target cell.

Another group of suitable targeting moieties are ligands wherein the target molecule is a receptor (for which the targeting moiety is a ligand) associated with the surface of the cell.

5 For many cell type-specific antigens protein ligands have been identified which bind with high specificity and/or affinity, e.g. cytokines binding to their cellular receptors. Most proteins can be labeled with the water-soluble vitamin biotin, using simple procedures. Most  
10 biotinylations are performed using succinimide esters of biotin. Binding to the protein proceeds through free amino groups, normally of lysyl residues. Biotinylated molecules are bound with exceptionally high affinity ( $K_a=10^{14}$ - $10^{15}$  M<sup>-1</sup>) by both Avidin and Streptavidin (Wilchek and Bayer,  
15 Immunol. Today 5(1984):39). Avidin is a 68 kD glycoprotein isolated from egg white and Streptavidin is a 60 kD protein from *Streptomyces avidinii*. Both molecules are homotetramers; each subunit contains a single biotin binding-site. The high affinity of their binding makes the  
20 biotin-avidin or biotin-streptavidin interactions essentially irreversible. A functional streptavidin gene has been cloned (Sano and Cantor, Proc. Natl. Acad. Sci. USA 87(1990):142) and a streptavidin mutant has been generated (Sano and Cantor, Proc. Natl. Acad. Sci. USA 92(1995):3180)  
25 with reduced biotin-binding affinity (approx.  $K_a=10^8$  M<sup>-1</sup>) providing specific and tight, yet reversible biotin-binding. As for Protein A and G, avidin and streptavidin do not intrinsically bind to mammalian cell surfaces or virus membranes or capsids. Expression of (parts of) these  
30 molecules on the surface of gene delivery vehicles will therefore also require the generation of hybrid molecules. Hybrids of streptavidin and heterologous proteins have been made and retained full biotin binding capacity (Sano and Cantor, Bio/Technol. 9(1991):1378; Sano et al., Proc. Natl.  
35 Acad. Sci. USA 89(1992):1534).

In yet another embodiment of the invention the member of the specific binding pair and its counterpart are both



peptides forming three-dimensional structures that can interact in solution. Peptides useful in this aspect of the invention include but are not restricted to dimerization motifs that are identified within proteins known to form dimers, such as the yeast transcription factor GCN4, the mammalian transcription factor C/EBP, and the nuclear transforming oncogene products fos, jun, and myc. Alternatively, synthetic peptides can be used that are designed on the basis of the knowledge of inter and intra protein interactions. The paradigm of peptide motifs known to dimerize is the coiled-coil structure, a subset of which is the so called leucine zipper (Landschultz et al., Science 240(1988):1759; O'Shea et al., Science 243(1989):538; O'Shea et al., Science 254(1991):539). Several proteins containing a leucine zipper motif can not only form homodimers but also heterodimers (Hai et al., Genes & Dev. 3 (1989):2083; Roman et al., Genes & Dev. 4(1990):1404). The leucine zipper dimerization region itself contains all of the structural information determining the dimerization specificity (Agre et al., Science 246(1989):922; Kouzarides and Ziff, Nature 340(1989):568; O'Shea et al., Science 245(1989):646). For the design of the member of the specific binding pair and its counterpart in the present invention it is preferred that the peptides used preferentially, if not exclusively, form heterodimers and not homodimers. Vinson et al (Genes & Dev. 7(1993):1047) have proposed and verified a rule for the rational design of leucine zipper motifs at preferentially form heterodimers. Therefore, it is preferred that the peptides used in this aspect of the invention are designed according to this so called "i+5 salt bridge" rule. Expression of a dimerizing peptide motif on the surface of the delivery vehicle as well as conjugation of its dimerizing counterpart to the targeting moiety will require the generation of hybrid molecules. Many different hybrid proteins with functional leucine zipper domains have already been produced, including dimeric antibodies based on scFv (Pack and Plückthun, Biochemistry 31(1992):1579), dimeric

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

antibodies based on monoclonal Fab' fragments (Kostelny et al., J. Immunol. 148(1992):1547), and a set of heterodimerizing proteins with leucine zippers, one linked to phagemid pIII coat protein and the other linked to the products from a cDNA library, that can be used for the production of a combinatorial phagemid cDNA library (Crameri and Suter, Gene 137(1994):69).

When the gene delivery vehicle is provided with a peptide structure capable of dimerization with a counterpart in solution it can be used in combination with any targeting moiety that can be coupled to said counterpart. Using standard techniques known in the art expression constructs can be made for insertion of foreign sequences resulting in the translation of fusion proteins containing a leucine zipper terminal domain. Said coupling may either be direct or via an intervening linker sequence separating the individual folding domains of said targeting moiety and counterpart. Linker sequences known in the art that provide the peptide main chain with conformational freedom, thereby separating individual folding domains, are usually rich in glycine residues. Examples of linker peptide sequences useful in the invention include but are not restricted to (Gly<sub>4</sub>-Ser)<sub>3</sub> (Batra et al., J. Biol. Chem., 265(1990):15198) and (Gly<sub>4</sub>-Thr)<sub>3</sub> (Bird et al., Science 242(1988):423). It is thus understood from the above that also in this aspect of the invention the number of targeting moieties that can be applied is almost limitless.

As stated above it is possible to target the gene delivery vehicles to many different cells using different conjugates (antibodies, biotinylated targeting moieties or targeting moieties conjugated to a peptide structure that can interact in solution). It is thus preferred to provide a kit of parts according to the invention comprising a multitude of different conjugates, comprising the same counterpart member but a number of different targeting moieties.

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

Although as stated above the skilled artisan will be able to apply the teaching of the present invention to other viruses than those exemplified herein, the exemplified viruses are preferred embodiments, because of the experience with the viruses in gene therapy concepts. Retroviruses are especially suitable for gene transfer into replicating cells. Foreign genes introduced by retroviruses become stable components of the genome of the target cell and its progeny. Adenoviruses, on the other hand, efficiently introduce foreign genetic material into non-dividing target cells. Adeno-associated virus is the only currently known non-pathogenic DNA-virus. It could provide an alternative for both retroviruses and adenoviruses in certain gene transfer applications. Thus it is preferred that the gene delivery vehicle is derived from an adenovirus, an adeno associated virus or a retrovirus.

Brief Description of the Drawings:

Figure 1 depicts the specific binding of biotinylated G-CSF to mouse fibroblasts expressing DG-CSFR on their membrane. FACS histograms of biotinylated G-CSF binding (log scale) are given. Solid lines represent measurement after incubation in biotinylated G-CSF, PE-Streptavidin, biotinylated goat-anti-Streptavidin, and PE-Streptavidin. Broken lines represent results of control incubations without biotinylated G-CSF. The percentage of cells showing specific binding of biotinylated G-CSF is shown in the upper right corner of each panel. Panel A: unmanipulated mouse fibroblasts, panel B: mouse fibroblasts transduced with LNCX-GRDcyt virus and selected with G418, panel C: mouse fibroblasts transduced with LNCX-GRDcyt virus, selected with G418, FACS sorted for GRDcyt expression and cloned by limiting dilution.

Figure 2 depicts the levels of hFcγRIa expression on the membrane of PA317 cells during in vitro culture for up to eight weeks. Unmanipulated PA317 cells, and two mixed

populations of transfected PA317 cells which were FACS sorted for hFcγRIa expression (PAFcR sorted-315 and PAFcR sorted-216) were stained with the anti-FcγRI MoAb 22 or with an irrelevant anti-KLH isotype control. The ratios between the median signal intensities after staining with MoAb 22 over the control are given.

Figure 3 depicts the structure of the two hybrid hFcγRIa/Mo-MuLV env molecules and their parental hFcγRIa and Mo-MuLV env proteins. The Mo-MuLV env peptides gp70 and p15E which are formed by proteolytic cleavage are shown separately and their physical connection on the precursor peptide is delineated by a broken line. Numbers above different segments indicate number of aminoacids. Asterisks indicate position of Cys-residues. Arrows represent primers used for PCR amplification. Areas in FcRenv-15 and FcRenv-70 indicated with striped boxes underneath were made by PCR amplification. Positions of relevant restriction enzyme cleavage sites are indicated. Mo-MuLV env sequences are shown in grey, hFcγRIa sequences in white. Black boxes represent non-coding Mo-MuLV sequences. S, signal peptide; Pro, proline-rich hinge-like region; TM, trans-membrane spanning domain; C, cytoplasmic domain; EC1, EC2, EC3, extracellular domains 1, 2, and 3, each forming one immunoglobulin-like loop.

Figure 4 shows *in vitro* immunoglobulin binding by hybrid hFcγRIa/Mo-MuLV env molecules FcRenv-15 and FcRenv-70. Positive and negative protein controls are wild type hFcγRIa (wt-FcR) and irrelevant firefly luciferase proteins, respectively. After *in vitro* coupled transcription and translation of the DNA sequences encoding these proteins, or a control reaction without DNA template (no DNA) in the presence of <sup>3</sup>H-labelled leucine, the formed proteins were precipitated with Sepharose beads loaded with IgG2a antibodies. The total reaction product (T), precipitated material (P) and precipitation supernatant (S) were separated through 10% SDS-polyacrylamide gel. The signal in

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

the gel was amplified and exposed to an X-ray film. Extended exposures are indicated with a - above the bands. The molecular weights (MW) of a marker protein mixture is indicated at the right side of the gel.

5

Detailed Description of the Invention:

The present invention provides a method to universally exploit a recombinant virus preparation for the targeted  
10 delivery of genetic material to many different cell types. To this end, the invention describes methods for the production of recombinant viruses carrying on their surface molecules (members of a specific binding pair) which can specifically interact with a set of secondary molecules  
15 (conjugates comprising the counterparts of the members of a specific binding pair). Said secondary molecules possess the ligand specificity for target cell-specific antigens. The indirect binding of the recombinant virus to the target cell via a single secondary molecule, which allows redirection of  
20 the same recombinant virus preparation to different target cells depending on the added secondary molecule, is a very important aspect of the invention. Useful secondary molecules to be employed in the invention include immunoglobulins or fragments thereof, chemically modified  
25 immunoglobulins or fragments thereof, chemically modified natural protein ligands for target cell antigens, fragments thereof, or recombinant derivatives thereof, and immunoglobulins, fragments thereof, recombinant derivatives thereof, natural protein ligands for target cell antigens,  
30 fragments thereof, or recombinant derivatives thereof that are coupled to a peptide motif that can interact with a specific counterpart in suspension. Biotinylation is the paradigm for the chemical modifications which could be employed. Typically, the specificity of the secondary  
35 molecules for the target cell antigen is high, and preferably the affinity of the secondary molecules for the target cell antigen is high. Example 1 describes the

specific interaction between a cytokine receptor molecule expressed on a mamalian cell surface and a biotinylated cytokine ligand. The invention is not restricted in the range of target cell antigens which could be employed to mediate targeted gene delivery. In principle, any target cell antigen for which a specific secondary molecule is available or can be made available is eligible for application of the invention. It is recognized, however, that not every target cell-specific molecule might serve as an internalization site for viruses bound to it. Furthermore, the efficiency of gene delivery may depend on the composition of the molecular bridge between the virus and the target cell-specific molecule (Etienne-Julan et al., J. Gen. Virol. 73(1992):3251). Hence, attempts to target gene delivery vehicles to specific cell types relying on methods in which a specific target cell ligand is expressed on the surface of the virus particle, either as a wild-type molecule or as a hybrid with a heterologous protein, can be extremely costly and time-consuming since certain chosen approaches may be destined to fail. The procedures disclosed in this invention, however, allow for comprehensive screening of useful target molecules by employing many different secondary molecules added to the same gene delivery vehicle preparation. Therefore, application of the invention will tremendously increase the chances for successful targeted gene delivery and allow optimization of the procedures at relatively low costs. In addition, standardization and validation of the gene delivery preparations will be simplified as compared to strategies using antibody-antibody or antibody-ligand complexes or methods relying on direct chemical modification of the virus envelope or capsid.

The binding of the secondary molecules to the recombinant viruses is mediated by substances which are exposed on the virus surface and have specific binding capacity for the secondary molecules. Preferably, said

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

substances bind to the secondary molecules with high affinity. Said substances are selected from or contain regions from proteins including but not restricted to Protein A, Protein G, FcR, anti-immunoglobulin scFv, Avidin, Streptavidin, proteins containing structural peptide domains that can specifically interact with a peptide domain counterpart, and recombinant derivatives from the above. Their exposure on the surface of recombinant viruses is envisaged in several ways, which are listed below:

10

A. for recombinant viruses with an envelope:

Recombinant retroviruses are described in the invention as the paradigm for recombinant enveloped viruses. The invention is, however, not restricted to retroviruses. The invention also applies to other enveloped viruses, such as e.g. rhabdoviruses or herpes viruses.

15

1. If the binding substance is itself a trans-membrane molecule, e.g. FcR, the complete wild-type binding substance can be expressed on the surface of the retrovirus producing cell line. Alternatively, retrovirus packaging constructs can be expressed in cells naturally expressing the binding substance. Whether these approaches will result in inclusion of the binding substance in the retrovirus particle depends at least in part on the nature of the substance. During the assembly of murine retroviruses at the cell surface cellular membrane molecules are merely excluded from the virus particle, although exceptions have been observed (Závada, J. Gen. Virol. 63(1982):15; Calafat et al., J. Gen. Virol. 64(1983):1241). In addition, exclusion of the binding substance from the virus membrane may be overcome by expressing it at very high density on the membrane of the retrovirus producing cell line (Suomalainen and Garoff, J. Virol. 68(1994):4879).

20

25

30

35

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

2. In the application of method no. 1 it is preferred that the trans-membrane binding substance lacks sequences which might elicit intracellular processes in the retrovirus producing cell line or intraviral processes in the virus particle. Such could perhaps occur upon binding of the secondary targeting molecule. Therefore, the binding substance can also be a modified derivative of the wild-type molecule. Modifications include any mutation preventing signal processing through the binding substance. The use of such truncated binding substances is furthermore preferred as to prevent interactions with cellular proteins in the retrovirus producing cell line. It has been suggested that interactions with cytoplasmic proteins might be partially responsible for the exclusion of host cell surface proteins from retrovirus particles (Young et al., Science 250(1990):1421).
3. Since the fusion process between an infective retrovirus particle and a target cell is assumed to be mediated by an as yet undefined region in the carboxy terminal envelope peptide it can be envisaged that target cell binding according to method no. 1 or 2 might be insufficient to obtain internalization of the gene delivery vehicle into the target cell. Therefore, it may be necessary to co-express the trans-membrane binding substances on the retrovirus surface with wild-type retroviral envelope molecules or segments thereof, which provide the retrovirus with fusion capability. Examples 2 and 3 describe the co-expression of hFcγRIa and wild-type retroviral envelope molecules on the surface of recombinant retrovirus producing cell lines and functional IgG Fc-binding by the exposed hFcγRIa molecules. It is preferred that the co-expressed envelope molecule is incapable of recognizing receptor molecules on cells present in the mixture of cells or in the living animal body subjected to the gene delivery preparation. This can be achieved by employing truncated envelope molecules lacking their receptor recognition site or wild-type



envelope molecules from a retrovirus species with a tropism restricted to cells not present in the mixture of cells or in the living animal body subjected to the gene delivery preparation.

- 5 4. Hybrid molecules can be made containing (segments of) a binding substance and (segments of) a heterologous protein which anchors the hybrid molecule in the membrane of the retrovirus producing cell line. This method also applies to binding substances which are not themselves trans-
- 10 membrane proteins. Molecules serving as a membrane anchor can be derived from natural trans-membrane proteins including but not restricted to cytokine or hormone receptors, cell adhesion or interaction molecules, complement receptors, and immunoglobulin receptors.
- 15 Alternatively, the hybrid molecule could comprise a membrane attachment region from a glycosylphosphatidylinositol-anchored protein. The invention is not restricted to a defined junction site between the heterologous molecules.
- 20 5. In the application of method no. 4 it is preferred that the membrane anchor molecule lacks sequences which might elicit intracellular processes in the retrovirus producing cell line or intraviral processes in the virus particle, or interact with cellular proteins in the retrovirus
- 25 producing cell line. It is, therefore, preferred that sequences conferring the above be deleted from the hybrid binding substance.
6. Because the retroviral envelope molecule is selectively incorporated in retroviral particles and is assumed to
- 30 mediate fusion between the virus and the target cell, it is preferred over methods nos. 4 and 5 to construct hybrid molecules containing the retroviral envelope peptide sequences responsible for the above processes and the peptide sequences from the binding substance responsible
- 35 for binding to the secondary target molecule. Also this method applies to binding substances which are not themselves trans-membrane proteins. The invention is not

restricted to a defined junction site between the heterologous molecules. Example 4 describes the construction of two such hybrid binding substances and Example 5 shows that said hybrid binding substances retain functional binding capacity for secondary target molecules.

7. Hybrid molecules can be made containing (segments of) a binding substance and (segments of) a heterologous protein which specifically interacts with extracellular regions of membrane molecules on the retrovirus producing cell surface. Preferably, the hybrid molecule interacts with membrane molecules which are specifically incorporated into retrovirus particles. Typically, these membrane molecules are (segments of) retroviral envelope proteins. This method also applies to binding substances which are not themselves trans-membrane proteins.
8. The expression of viral membrane proteins on the cell surface requires correct folding and assembly in the endoplasmatic reticulum (ER) of the virus producing cell line (reviewed by Doms et al., Virology 193(1993):545). Retroviral envelope molecules assemble into dimers, trimers, or tetramers, depending on the retrovirus species. Incorrect oligomerization may prevent proper transport to the Golgi apparatus, leading to degradation of the molecule. It can be anticipated that this would be the faith of some hybrid substances produced according to methods nos. 4 to 7. This could possibly be prevented by co-expressing the hybrid substance with wild-type envelope molecules or segments thereof. The formation of hetero-oligomers might ensure correct processing in the ER. The preferred requirements for co-expressed envelope molecules described under no. 3 also apply here.

The principles of methods A 1-8 also apply to pseudotyped viruses in which the envelope glycoproteins from one virus species are carried on the membrane of a heterologous virus species. Such phenotypic mixing of

envelope glycoproteins occurs between a variety of different virus families (reviewed by Závada, J. gen. Virol. 63(1982):15). The invention also applies to enveloped viruses with hybrid envelope proteins, comprising fragments of related or even unrelated viruses. It has, e.g., been shown possible to generate infectious recombinant retroviruses carrying hybrid influenza hemagglutinin/retrovirus envelope proteins (Dong et al., J. Virol. 66(1992):7374).

B. for viruses with a capsid:

The invention is described for, but not restricted to recombinant adenoviruses and adeno-associated viruses. Hybrid molecules can be made containing (segments of) a binding substance and (segments of) virus capsid proteins. For adenoviruses, it is preferred to locate the binding substance in the head domain of the fiber protein. This domain carries the receptor recognition site and it has been shown possible to change the adenovirus receptor specificity by manipulating this segment (Stevenson et al., J. Virol. 69(1995):2850). For certain hybrid molecules it may be necessary to co-express the hybrid molecule with wild-type fiber protein to ensure proper fiber trimerization and association with the penton base complex. Alternatively, other capsid proteins of adenovirus may be used to incorporate the binding substance. E.g., Curiel et al. (Hum. Gene Ther. 3(1992):147) have generated a chimeric serotype 5 adenovirus that contains a *Mycoplasma pneumoniae* P1 protein epitope as part of the hexon protein.

It is furthermore preferred that the natural adenovirus receptor binding sequences be deleted from the hybrid molecule. Hybrid AAV capsid proteins with altered binding specificity are preferably made using (segments of) VP3, since the AAV capsid mainly comprises of this molecule. Alternatively, VP1 or VP2 could be used. The AAV viral proteins allow insertions which are exposed on the capsid

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

surface (unpublished results from the group of Dr. R. Samulski, Univ. of North Carolina, Chapel Hill, NC). For certain hybrid molecules it will be necessary to co-express the hybrid molecule with wild-type capsid protein. The invention is not restricted to specific junction sites between the heterologous proteins.

Since many virus capsids self-assemble independent of the presence of viral DNA, empty pseudocapsids can be generated *in vitro*. Assembly of empty AAV-like particles does not even require all three capsid proteins (Ruffing et al., J. Virol. 66(1992):6922). Thus, there is significant freedom to design novel pseudocapsids. Empty virus-like particles can be used to package and transfer exogenous non-viral DNA. This has been shown for Polyoma Virus pseudocapsids (Forstová et al., Hum. Gene Ther. 6(1995):297). Furthermore, exogenous non-viral DNA can be chemically coupled onto the exterior of virus capsids (Curiel et al., Hum. Gene Ther. 3(1992):147; Cotten et al., Proc. Natl. Acad. Sci. USA 89(1992):6099). The invention emphatically also applies to the latter two approaches and their combination, i.e. chemical coupling of exogenous DNA to the outer surface of empty capsids.

**EXAMPLES****Example 1**

- 5 Specific binding of biotinylated G-CSF to G-CSFR molecules  
expressed on the surface of mouse fibroblasts.

To express receptors for granulocyte-colony stimulating factor (G-CSF) on the cell membrane of murine fibroblasts,  
10 PA317 cells (Miller and Buttimore, Mol. Cell. Biol. 6(1986):2895) were transduced with ecotropic LNCX/GRDcyt virus. The construct pLNCX/GRDcyt (generated in the laboratory of Dr. I. Touw, Erasmus University Rotterdam, The Netherlands) was made by inserting the cDNA sequence for a  
15 mutant G-CSF receptor (DG-CSFR) into pLNCX (Miller and Rosman, BioTechniques 7(1989):980). DG-CSFR differs from the wild-type molecule by an almost complete deletion of the intracellular domain, rendering the receptor incapable of signal-transduction. The extracellular and trans-membrane  
20 domains of DG-CSFR are of wild-type structure. DG-CSFR exhibits, therefore, normal binding capacity for its cytokine ligand. Transduced PA317 cells were isolated by selecting for resistance to 1 mg/ml G418 (Geneticin; Gibco, Paisley, UK), conferred by expression of the *neo<sup>r</sup>* gene in  
25 LNCX/GRDcyt. Resistant PA317 cells were incubated with biotinylated G-CSF. Bound G-CSF was visualised on a Fluorescence Activated Cell Sorter (FACS) after incubation with PE-conjugated Streptavidin (Molecular Probes, Eugene, OR), biotinylated goat-anti-Streptavidin antibodies (Vector  
30 Laboratories, Burlingame, CA), and a second incubation with PE-Streptavidin. As can be seen in figure 1, untransduced PA317 cells did not bind any biotinylated G-CSF, whereas approximately 27% of the transduced cells bound G-CSF. After FACS sorting of positive cells and cloning by limiting  
35 dilution, a clone was isolated which strongly bound biotinylated G-CSF (figure 1C). This example shows that a

biotinylated cytokine specifically binds to its natural receptor expressed on the membrane of a mammalian cell line.

#### Example 2

#### 5 Generation of recombinant retrovirus packaging cell lines expressing hFcγRIa proteins on their cell membrane.

For expression of functional hFcγRIa on the surface of retrovirus packaging cell lines the construct pRc/CMV-hFcγRIa was used. This construct was generated by inserting a 1.3 kb HindIII-NotI fragment including the full-length p135 hFcγRIa-cDNA sequence (Allen and Seed, Science 243(1989):378; GenBank accession number M21090) from clone CDM into the polylinker of pRc/CMV. Twenty microgram pRc/CMV-hFcγRIa was transfected onto the ecotropic packaging cell line GP+E-86 (Markowitz et al., J. Virol. 62(1988):1120) and onto the amphotropic packaging cell line PA317, according to the method described by Chen and Okayama (Mol. Cell. Biol. 7(1987):2745). Transfectants were selected for resistance to 1 mg/ml G418, conferred by expression of the SV40pr-neo<sup>r</sup>-p(A) cassette on the pRc/CMV-hFcγRIa construct. Pools of resistant cells were analysed for hFcγRIa expression on a FACS, after staining with the anti-FcγRI monoclonal antibody (MoAb) 22 (Guyre et al., J. Immunol. 143(1989):1650) and FITC-conjugated goat-anti-mouse IgG antibodies (GaM-FITC; Becton Dickinson Immunocytometry Systems, San Jose, CA [BDIS]). Table 1 shows that only few G418-resistant GP+E-86 cells (E86FcR) and G418-resistant PA317 cells (PAFcR) were found to express FcγRI on their surface. FcγRI-expressing cells were sorted using the FACS and expanded. After two weeks of culture, the sorted populations were analysed for FcγRI-expression as before. As can be seen in table 1, the sorting procedure resulted in significantly enriched FcγRI-expressing E86FcR and PAFcR cell populations.

Table 1. FcγRI Expression on pRc/CMV-hFcγRIa Transfected and FACS Sorted Ecotropic and Amphotropic Recombinant Retrovirus Packaging Cell Lines.

Packaging Cells	FcRI Positive Cells (%) <sup>1</sup>	Average FcγRI Expression <sup>2</sup>
GP+E-86	0.1	N.D.
PA317	0.2	0.98
E86FcR	0.9 <sup>3</sup>	N.D.
E86FcR sorted	16.4	1.16
PAFcR	7.6 <sup>3</sup>	N.D.
PAFcR sorted-315 <sup>4</sup>	10.5	1.43
PAFcR sorted-216 <sup>4</sup>	20.5	2.37

5   <sup>1</sup> Percentage cells exhibiting FITC fluorescence following incubation with MoAb 22 and GaM-FITC. Controls incubated with GaM-FITC only were set to ≤ 0.1 %.

2 Relative median FITC fluorescence after incubation with MoAb 22 and GaM-FITC as compared to incubation with  
10 irrelevant anti-KLH isotype control MoAb (BDIS) and GaM-FITC.

<sup>3</sup> Average of two independent measurements.

<sup>4</sup> PAFcR cells were subjected to two independent sorting experiments.

15 N.D., not done.

To study whether hFcγRIa remained stably expressed on the packaging cell surface in the absence of g-chain co-expression, two sorted cell populations expressing different levels of hFcγRIa as deduced from their median FITC-signals after staining with MoAb 22 as compared to staining with irrelevant isotype controls, were cultured for up to eight weeks in culture medium containing G418. Regular reanalysis showed rather stable median fluorescence signals (approx. 1.5 and 2.3 for the two sorted populations, as compared to 1.0 for untransfected PA317 cells; figure 2) throughout the observation period.

Immunophenotypic analysis of wild-type hFcγRIa revealed three distinct epitopes, one of which comprises the Fc-binding domain (Guyre et al., J. Immunol. 143(1989):1650). One of the sorted cell populations was analysed using the directly FITC-stained MoAbs 22 and 32.2 (Medarex, Annandale, NJ), which each define one of the two non-Fc-binding epitopes. Table 2 shows that both MoAbs recognized the hFcγRIa molecule exposed on the cell surface. The increase in fluorescence which was observed when both MoAbs were combined demonstrate that they bind to distinct epitopes on the packaging cell surface. Pre-incubation of the cells with irrelevant unstained mouse immunoglobulins of IgG1 or IgG2a isotype (BDIS) did not influence the binding of MoAbs 22 and 32.2 (not shown). These results demonstrate that hFcγRIa of correct immunophenotypic structure was expressed on the cell surface of a retrovirus packaging cell line.



Table 2. Immunophenotype Analysis of Amphotropic Packaging Cells Expressing hFcγRIa.

FITC-labeled MoAb	% Positive Cells	Mean Fluorescence Intensity (a.u.)
anti-KLH (irrelevant)	0.9 <sup>1</sup>	15.0
anti-hFcγRIa MoAb 22	13.0	64.6
anti-hFcγRIa MoAb 32.2	8.1	46.3
anti-hFcγRIa MoAbs 22 + 32.2	21.1	105.8

PAFcr sorted-216 cells were incubated with the indicated FITC-labeled MoAbs and analysed on a FACS. <sup>1</sup> The threshold for a positive score was set at ≥ 99 % negative cells on the anti-KLH-FITC stained sample.

Ecotropic and amphotropic hFcγRIa-expressing populations were cloned by limiting dilution. Twelve individual E86Fcr and eleven individual PAFcr clones were expanded and analysed for hFcγRIa-expression. The two clones expressing the highest level of hFcγRIa from each population were used to investigate whether hFcγRIa-expression on the cell surface of packaging cells influences the retrovirus production. To this end, cell-free supernatant was harvested from these four clones as well as from the parental packaging cell lines under standard conditions (72 hours at 32°C in 10 ml alpha-modified DMEM with 10% BCS per 80 cm<sup>2</sup> confluent cell monolayer). The supernatants were analysed for reverse transcriptase activity (Goff et al., J. Virol. 38(1981):339), in a two-fold dilution titration. Comparison of the radiation intensity of incorporated <sup>32</sup>P-dTTPs by exposure to X-ray film revealed that expression of hFcγRIa did not influence the shedding of virus particles into the culture medium more than a factor 2.3 (table 3).

Table 3. Retrovirus Particle Production by Packaging Cell Lines Expressing hFcγRIa.

Packaging Cell Line	FcγRI Expression <sup>1</sup>	Retrovirus Production <sup>2</sup>
PA317	1.2	(1)
PAFcr-2	2.3	2.0
PAFcr-8	2.7	2.0
GP+E-86	N.D.	1.8
E86Fcr-3	2.8	0.8
E86Fcr-14	2.6	1.3

<sup>1</sup> Relative median fluorescence signal after staining with FITC-labeled MoAbs 22+32.2 as compared to anti-KLH-FITC.

5 <sup>2</sup> The RT-activity in the culture supernatant is given relative to the RT-activity shed by PA317 cells.

N.D., not done.

### Example 3

10 Immunoglobulin binding by hFcγRIa proteins expressed on the cell membrane of recombinant retrovirus packaging cell lines.

To test whether hFcγRIa expressed on the surface of  
 15 retrovirus packaging cells is capable of specific immunoglobulin-Fc binding, PAFcr sorted-315 cells were incubated with irrelevant mouse immunoglobulins of IgG1 or IgG2a isotype. FcγRI molecules should bind IgG2a molecules with high affinity and IgG1 molecules with low affinity. As  
 20 can be seen in table 4, approximately one third of the sorted cells were expressing low amounts of hFcγRIa, as indicated by MoAb22-staining with low relative median signal. All hFcγRIa molecules on PAFcr cells also bound

mouse IgG2a (similar percentage positive cells and signal intensity), whereas mouse IgG1 was hardly bound. On human primary monocytes, which were used as a positive control, hFcγRIa molecules were strongly expressed and specifically bound mouse IgG2a immunoglobulins. The capacity of hFcγRIa molecules on monocytes to bind immunoglobulins, however, was lower than measured on pRc/CMV-hFcγRIa transfected packaging cells (55% of MoAb22-positive monocytes bound IgG2a, with a much lower signal intensity). In conclusion, all hFcγRIa molecules characterised by MoAb22-binding expressed on the surface of PAFcR cells exhibit a correct structure for specific high affinity binding of immunoglobulins.

Table 4. Immunoglobulin-Fc Binding by FcγRI Expressed on PA317 Cells.

Primary MoAb	Human Monocytes		PA317		PAFcr sorted-315	
	% Pos.	Rel. Med <sup>1</sup>	% Pos.	Rel. Med.	% Pos.	Rel. Med.
none	0.8	(1)	0.9	(1)	0.8	(1)
irr-IgG1	0.7	1.84	0.6	0.96	6.2	1.15
irr-IgG2a	35.1	3.69	0.4	0.95	30.6	1.61
MoAb22	64.2	8.82	0.5	0.94	33.0	1.65

Human primary monocytes, PA317 cells, or a pool of pRc/CMV-hFcγRIa transfected PA317 cells partially expressing hFcγRIa (PAFcr sorted-315) were first incubated with MoAb22 (IgG1 isotype), anti-KLH IgG1, anti-KLH IgG2a, or PBS/1% BSA ("none"), followed by an incubation with GaM-FITC. The percentage positive cells and the median fluorescence intensities were determined on a FACS.

<sup>1</sup> Relative median fluorescence intensity of cells incubated with indicated primary MoAb and GaM-FITC as compared to cells incubated with GaM-FITC alone.

5 Example 4

Construction of hybrid hFcγRIa/Mo-MuLV envelope genes.

For the construction of hybrid molecules with immunoglobulin-binding properties of hFcγRIa and virus-  
incorporation and membrane-fusion properties of envelope glycoprotein, pRc/CMV-hFcγRIa (see above) was used in  
10 combination with Mo-MuLV env sequences (Shinnick et al., Nature 293(1981):543). Two different hybrid hFcγRIa/Mo-MuLV-env constructs were made (figure 3). In both constructs, 3'  
15 hFcγRIa sequences in the pRc/CMV-hFcγRIa construct were replaced by 3' Mo-MuLV env sequences. Both constructs contain p135 sequences from the leader sequence to the  
HinPII-site at position 849-852. This fragment comprises the three extracellular domains of hFcγRIa, excluding the 61  
20 carboxy terminal nucleotides. In hybrid construct pRc/CMV-FcRenv-15, the p135-fragment is coupled to p15E env sequences encoded by a fragment starting from the HpaI-site  
at position 7195-7200 of Mo-MuLV until position 7823 in the IR sequence of the 3'LTR. To this end, the HpaI-site was  
25 converted into a HinPII-site by PCR, and a NotI-site was introduced in the 3'LTR by PCR. An internal SpeI-PvuII fragment (nt 7487-7747) was exchanged for cloned sequences.  
The correct sequence of the hFcγRIa/Mo-MuLV-env junction and of all sequences generated by PCR was confirmed by  
30 sequencing. Hybrid gene FcRenv-15 encodes an in-frame fusion protein comprising the extracellular domains of hFcγRIa until an Arg-residue 12 aminoacids downstream from the 3'  
Cys-residue forming the third immunoglobulin-like extracellular loop and all but the five most 5' p15E  
35 aminoacids. To generate hybrid construct pRc/CMV-FcRenv-70, the HinPII-site from the 5' p135-fragment was made blunt-end and was ligated to a BamHI (position 6537-6542)-PvuII

(position 7745-7750) fragment of Mo-MuLV, after the BamHI-site was made blunt-end, and the 3' PvuII-NotI PCR-fragment from pRc/CMV-FcRenv-15. Ligation of the blunt-end HinPII and BamHI-sites restored the BamHI-site. The correct structure of the hFcgRIa/Mo-MuLV-env junction was confirmed by sequencing. Hybrid gene FcRenv-70 comprises the same hFcgRIa-fragment as described for pRc/CMV-FcRenv-15 coupled to the Mo-MuLV env sequences starting from an Ile-residue at the BamHI-site in the amino terminal domain of gp70 and including the complete proline-rich hinge-like region and carboxy terminal domain of gp70 and the complete p15E peptide.

On July 24, 1996, the applicants have deposited with the European Collection of Cell Cultures (ECACC), Salisbury, Wiltshire, U.K., the plasmid pRc/CMV-FcRenv-15, given ECACC accession number P96072515, and plasmid pRc/CMV-FcRenv-70, given ECACC accession number P96072514. These deposits were made under the provisions of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the purposes of patent procedure and the regulations thereunder.

#### Example 5

Hybrid hFcgRIa/Mo-MuLV envelope genes FcRenv-15 and FcRenv-70 encode proteins of correct molecular weight that have immunoglobulin binding capacity

To test whether the fusion genes FcRenv-15 and FcRenv-70 encode functional hybrid molecules with immunoglobulin binding properties, the pRc/CMV-FcRenv-15 and pRc/CMV-FcRenv-70 constructs (Example 4) were used as templates for coupled in vitro transcription and translation. Transcription was initiated at the T7 promoter upstream of the gene insertion in the pRc/CMV vector. One microgram pRc/CMV-FcRenv-15 or pRc/CMV-FcRenv-70 DNA was added to a 25 microliter leucine-free T7 TnT coupled reticulocyte lysate reaction mixture (Promega, Madison, WI)

with 20 units RNasin ribonuclease inhibitor (Promega) and 20 microCi <sup>3</sup>H-leucine (Amersham, Buckinghamshire, UK). The reaction was allowed to proceed for 120 minutes at 30°C according to the guidelines provided by the manufacturer.

5 Control reactions were performed without DNA template, with an irrelevant control DNA template (firefly luciferase control DNA provided with the T7 TnT coupled reticulocyte lysate system), and with positive control pRc/CMV-hFcγRIa encoding wild-type hFcγRIa. To 5 microliter of the reaction

10 mixture, 20 microliter SDS-PAGE loading buffer (to a final concentration of 62.5 mM Tris pH 6.8, 10% glycerol, 2% beta-mercaptoethanol, 0.5% SDS, 0.0025% bromo phenol blue) was added. The remaining 20 microliter was diluted with 180 microliters of precipitation buffer (10 mM Tris-HCl pH 7.5,

15 150 mM NaCl, 5 mM EDTA, 1% Triton-100 with protease inhibitors PMSF at 0.1 mM, aprotinin at 10 microgram/ml, pepstatin at 5 microgram/ml, trypsin inhibitor at 10 microgram/ml, and leupeptin at 10 microgram/ml). Next, 10 microliter KLH-aKLH-IgG2a-Sepharose slurry was added and

20 mixed by tilted rotation for 90 minutes at room temperature. Said slurry was prepared according to the following procedure. First, 16.4 mg Keyhole Limpet Hemocyanin (KLH; Sigma, St. Louis, Mo) was coupled at 1.8 mg/ml to 1 gram CNBr-activated Sepharose 4B (Pharmacia Biotech, Uppsala, Sweden) according to the manufacturers instructions. The

25 KLH-Sepharose product was stored in PBS with 0.002% sodium azide at 4°C. Next, KLH-Sepharose was mixed 1:1 with 50 microgram/ml anti-KLH-IgG2a monoclonal antibody (Beckton-Dickinson Immunocytometry Systems, San Jose, CA) and

30 incubated for 30 minutes at room temperature. Finally, the KLH-aKLH-IgG2a-Sepharose beads were washed two times in 20 volumes PBS. This procedure should result in Sepharose beads coated with immunoglobulins of the IgG2a isotype that have their Fc domain projecting outwards. After incubation

35 of the *in vitro* transcribed and translated reaction product with KLH-aKLH-IgG2a-Sepharose, the KLH-aKLH-IgG2a-Sepharose beads were spun down and the precipitation supernatant was

removed. To 5 microliter precipitation supernatant  
20 microliter SDS-PAGE loading buffer was added as above.  
The pelleted KLH-aKLH-IgG2a-Sepharose beads were washed 6-  
times in 1 ml precipitation buffer. After a final spin,  
5 25 microliter SDS-PAGE loading buffer was added to the  
pellet. The above described procedure resulted in three  
different samples of each coupled *in vitro* transcription-  
translation reaction, i.e., the total reaction product, the  
immunoglobulin bound precipitated product, and the remaining  
10 material in the precipitation supernatant. Ten microliters  
of these samples were heated at 85°C for 2 minutes and  
subsequently separated by SDS-PAGE through a 10% gel  
according to standard procedures (Laemmli, Nature  
227(1970):680). After separation, the proteins in the gel  
15 were fixed in water/isopropanol/acetic acid (65%/25%/10%  
vol.) for 30 minutes at room temperature and thereafter  
incubated for 15 minutes at room temperature in Amplify  
(Amersham) for fluorographic enhancement of the signal.  
Finally, the gel was dried for 90 minutes at 80°C under  
20 vacuum and exposed to X-ray film with an enhancing screen.  
Figure no. 5 shows the result of this experiment. As can be  
seen, the constructs with genes encoding wild-type hFcγRIa  
or hFcγRIa/Mo-MuLV envelope fusion molecules all expressed  
proteins exhibiting a migration pattern corresponding to  
25 their predicted molecular weights, i.e., 42.5 kD for  
hFcγRIa, 52 kD for FcRenv-15, and 75,6 kD for FcRenv-70. The  
latter gene furthermore encodes a smaller protein product.  
This molecule perhaps represents the approximately 60 kD  
proteolytically processed protein after release of the p15E  
30 peptide. As can furthermore be seen in Figure no. 5, all  
these protein products were precipitated by KLH-aKLH-IgG2a-  
Sepharose beads, in contrast to the irrelevant control  
protein luciferase, demonstrating that the immunoglobulin  
binding property of wild-type hFcγRIa is retained after  
35 fusion of the three extracellular domains of hFcγRIa to Mo-  
MuLV envelope sequences.

PCT 0489

**CLAIMS**

1. A method for delivering genetic material to a target cell, comprising  
preparing a gene delivery vehicle comprising a molecule encoding a gene of interest, a capsid or an envelope derived  
5 from a virus and a first member of a specific binding pair,  
preparing a conjugate comprising a part which is a complementary member of said specific binding pair and a part which has affinity for a molecule associated with the surface of the target cell,  
10 contacting the gene delivery vehicle and the conjugate, so that the members of the specific binding pair can bind and delivering the complex of conjugate and gene delivery vehicle to said target cell.
2. A method according to claim 1, whereby said first  
15 member of a specific binding pair has no specific affinity for molecules associated with the surface of the target cell.
3. A gene delivery vehicle for use in a method according to claim 1 or 2, comprising  
20 an expressible recombinant nucleic acid molecule encoding a gene product of interest,  
a capsid or an envelope derived from a virus,  
and a member of a specific binding pair, the counterpart of which is not directly associated with the  
25 surface of the target cell.
4. A gene delivery vehicle according to claim 3, whereby the envelope or the capsid material is incapable of specifically binding to a target cell.
5. A conjugate for use in a method according to claim 1 or  
30 2 targeting a gene delivery vehicle according to claim 3 or 4 to a target cell comprising the counterpart of the member of a specific binding pair coupled to a targeting moiety being capable of binding to a target molecule associated with the surface of a target cell.

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**



6. A targeting gene delivery vehicle complex, comprising a gene delivery vehicle according to claim 3 or 4, coupled to a conjugate according to claim 5.
7. A kit of parts for delivering genetic material to a target cell, comprising a gene delivery vehicle according to any one of claims 3-4 and at least one conjugate of the counterpart member of the specific binding pair coupled to a targeting moiety, said targeting moiety being capable of binding to a target molecule associated with the surface of a target cell.
8. A kit of parts according to claim 7, wherein the member of a specific binding pair is an immunoglobulin binding moiety and its counterpart is an immunoglobulin or a derivative or a fragment thereof.
9. A kit of parts according to claim 8, whereby the immunoglobulin binding moiety is capable of binding to a constant region of an immunoglobulin.
10. A kit of parts according to claim 9, whereby the immunoglobulin binding moiety is protein A, protein G, or an Fc receptor.
11. A kit of parts according to claim 7 wherein the member of a specific binding pair is biotin and its counterpart is avidin or streptavidin.
12. A kit of parts according to claim 7 wherein the member of a specific binding pair is avidin or streptavidin and its counterpart is biotin.
13. A kit of parts according to anyone of claims 7-12, wherein the targeting moiety is an antibody or a fragment or a derivative thereof, recognizing the target molecule associated with the surface of the target cell.
14. A kit of parts according to anyone of claims 7-12, wherein the target molecule is a receptor for which the targeting moiety is a ligand.
15. A kit of parts according to anyone of claims 7-14, wherein the gene delivery vehicle is derived from an adenovirus, an adeno associated virus or a retrovirus.

**RECTIFIED SHEET (RULE 91)**  
**ISA/EP**

16. A kit of parts according to anyone of claims 7-15, comprising a multitude of different conjugates, comprising the same counterpart member but a number of different targeting moieties.

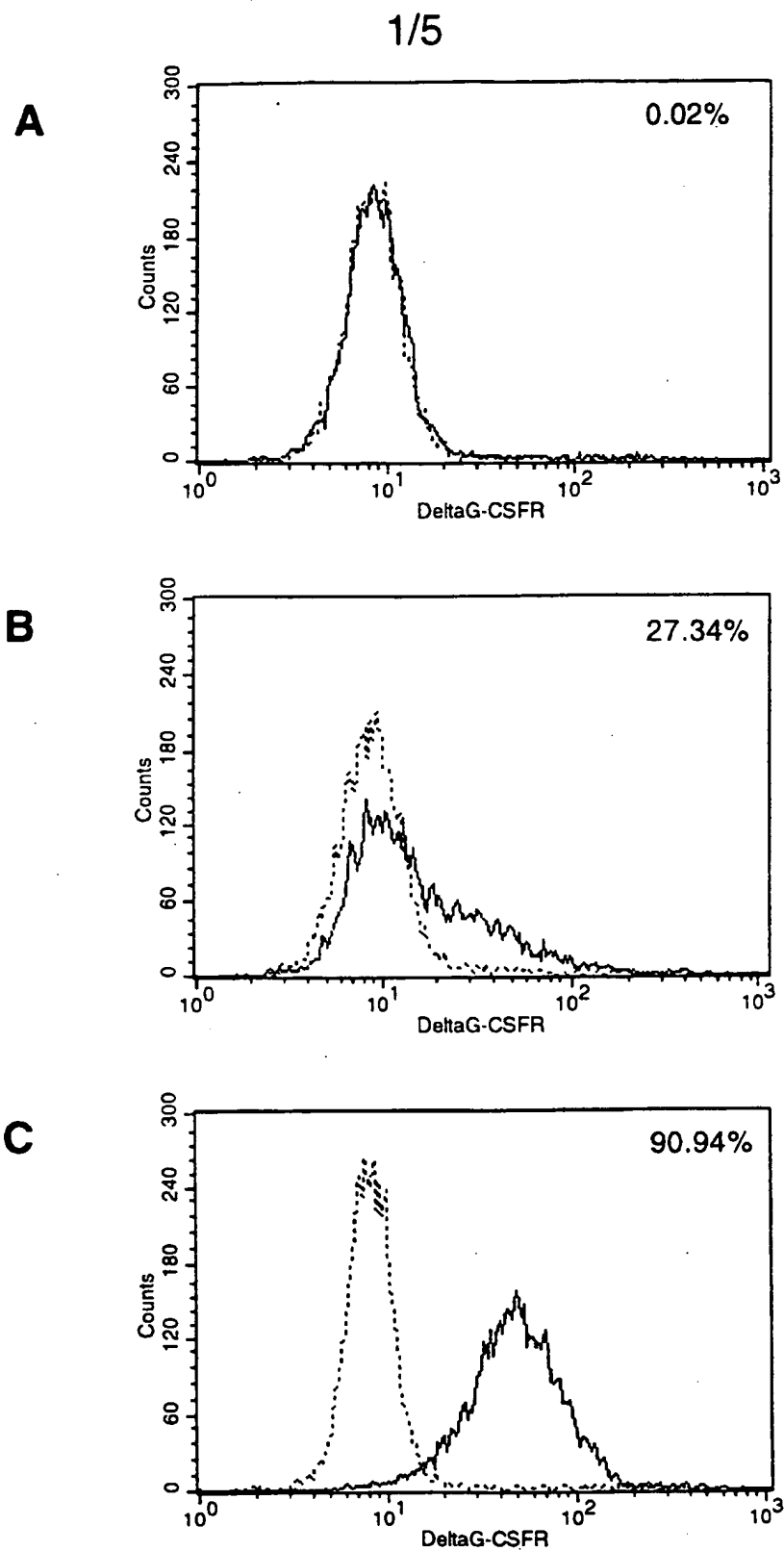


FIG. 1

SUBSTITUTE SHEET (RULE 26)

2/5

Stability of hFcγRIa expression on PA317 cells.

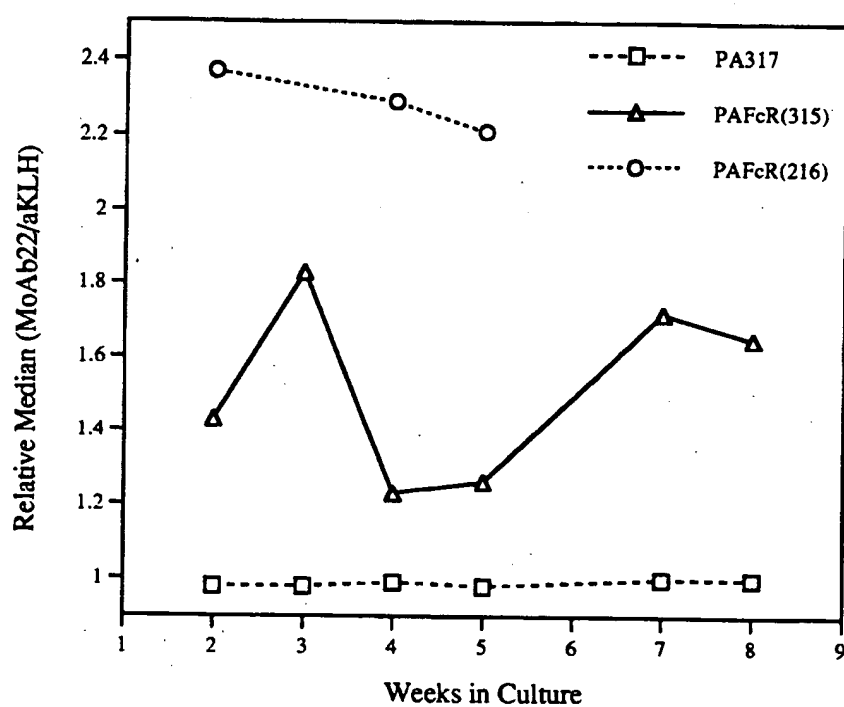


FIG. 2

SUBSTITUTE SHEET (RULE 26)

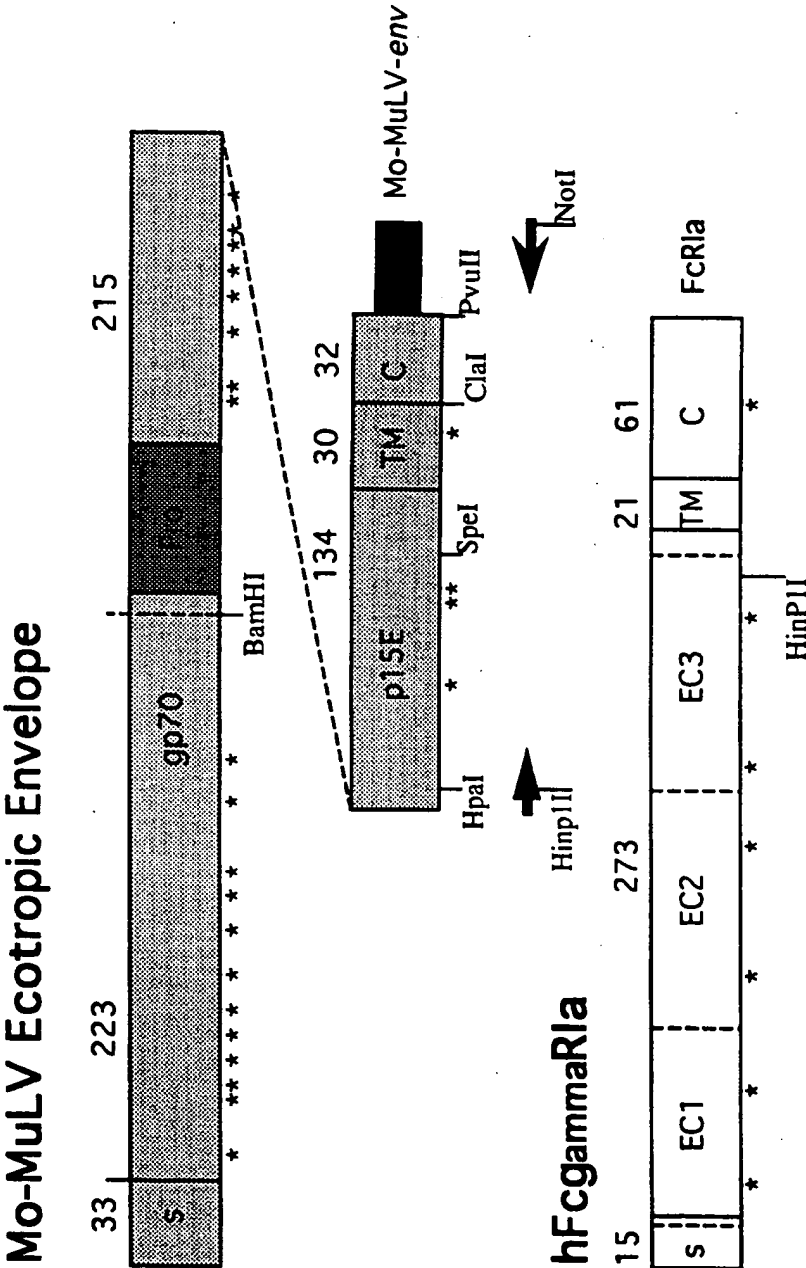


FIG. 3(1/2)

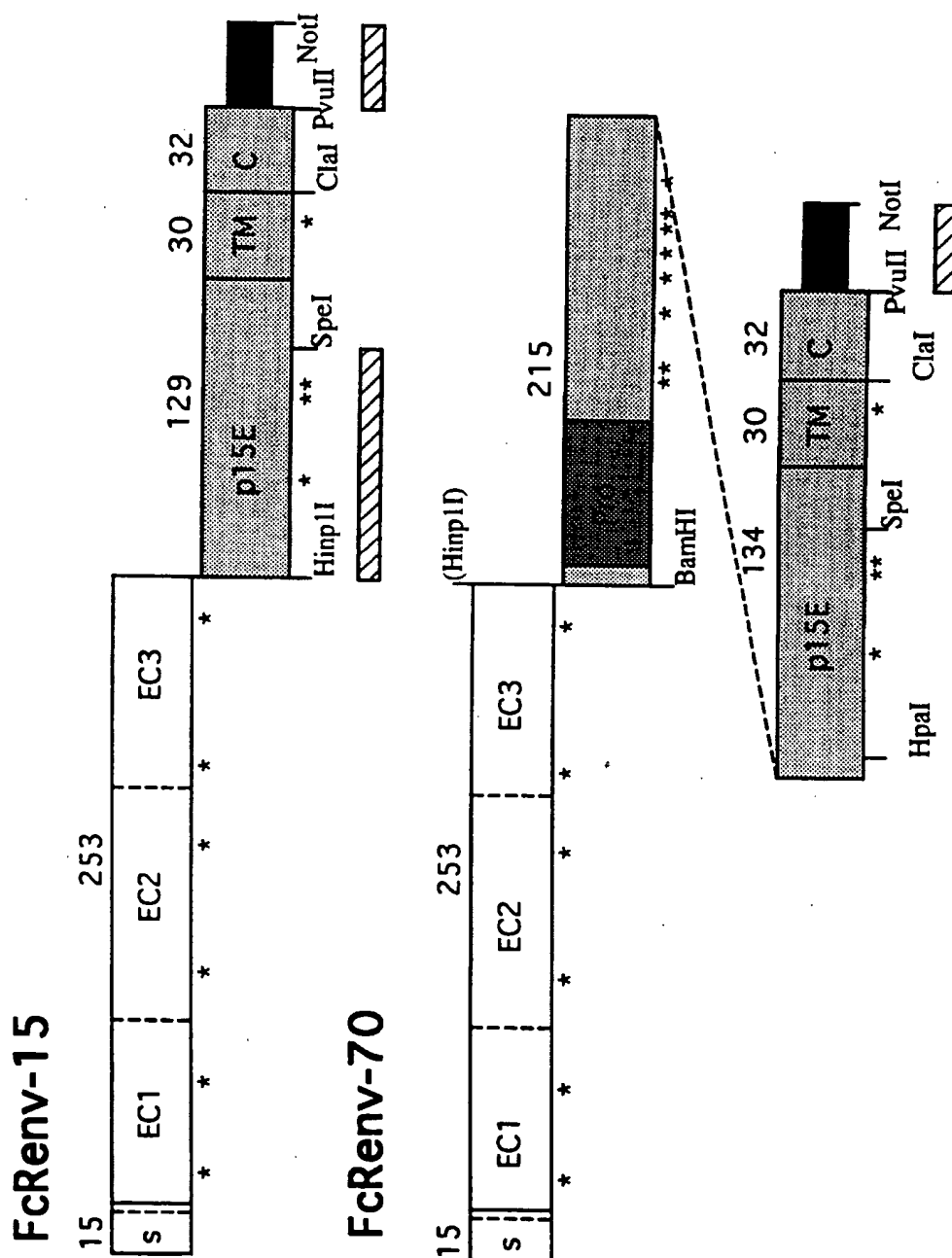
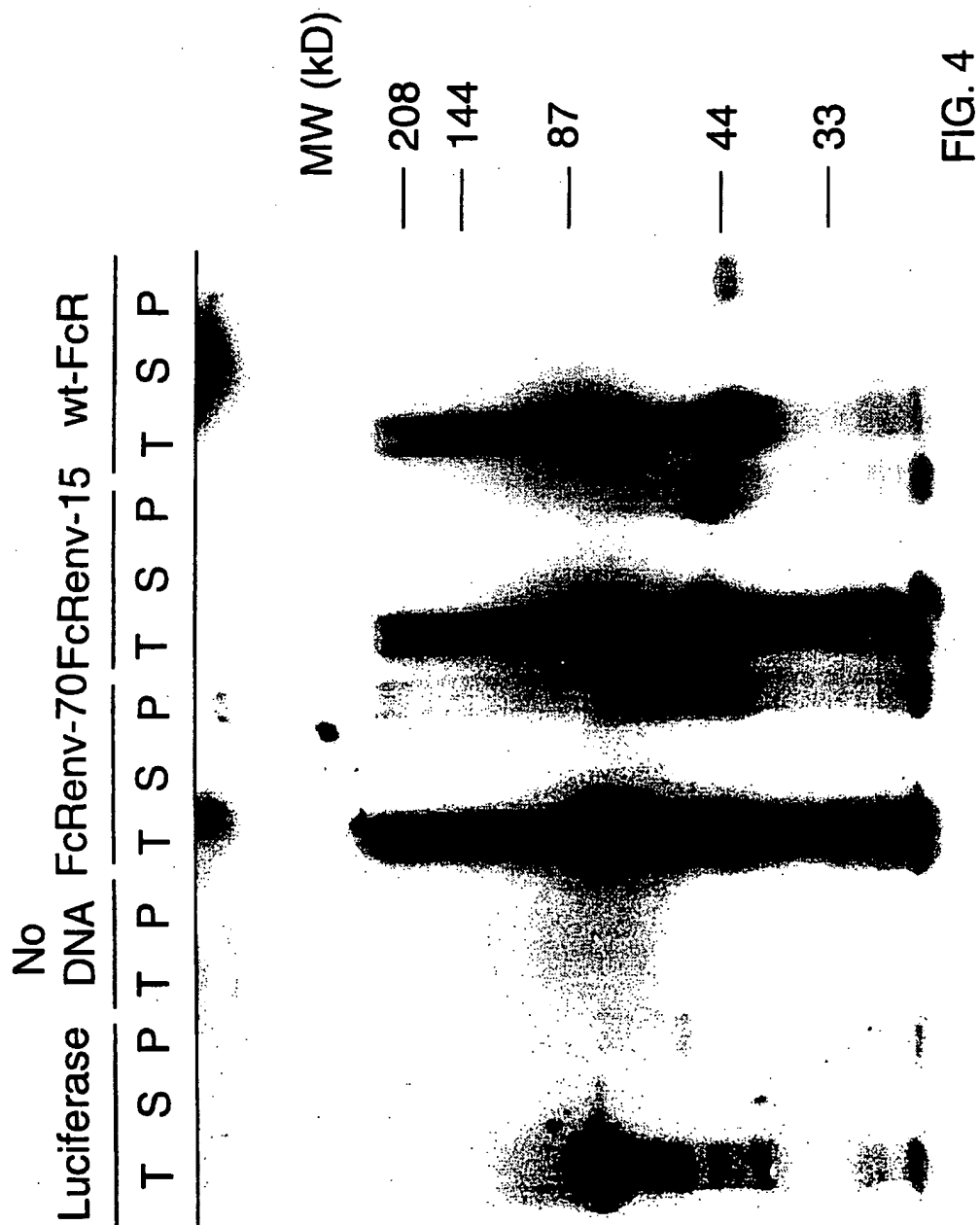


FIG. 3(2/2)



SUBSTITUTE SHEET (RULE 26)

# INTERNATIONAL SEARCH REPORT

International Application No  
PCT/NL 96/00302

<b>A. CLASSIFICATION OF SUBJECT MATTER</b> IPC 6 C12N15/87 A61K48/00 A61K47/48 C12Q1/70 //C12N15/86		
According to International Patent Classification (IPC) or to both national classification and IPC		
<b>B. FIELDS SEARCHED</b> Minimum documentation searched (classification system followed by classification symbols) IPC 6 C12N A61K		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practical, search terms used)		
<b>C. DOCUMENTS CONSIDERED TO BE RELEVANT</b>		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	VIROLOGY, vol. 198, no. 2, 1994, pages 577-585, XP002016471 CURIEL ET AL.: "Efficient foreign gene expression in Epstein-Barr Virus-transformed human B-cells" see the whole document ---	1-3,5-15
X	FR,A,2 649 119 (CENTRE NATIONAL DE LA RECHERCHE SCIENTIFIQUE) 4 January 1991 see page 1 - page 4 --- -/--	1-3,5-7, 13-15
<input checked="" type="checkbox"/> Further documents are listed in the continuation of box C. <input checked="" type="checkbox"/> Patent family members are listed in annex.		
* Special categories of cited documents : "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "A" document member of the same patent family		
Date of the actual completion of the international search  22 October 1996		Date of mailing of the international search report  05.11.96
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fac (+31-70) 340-3016		Authorized officer  Gac, G

Form PCT/ISA/210 (second sheet) (July 1992)



## INTERNATIONAL SEARCH REPORT

 Int. Patent Application No  
 PCT/NL 96/00302

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	J. CELL. BIOCHEM., vol. 17d, no. suppl., 1993, page 129 XP000608338 ZATLOUKAL ET AL.: "Receptor-mediated cytokine gene delivery to tumor cells for generation of cancer vaccines" * abstract nr NZ 522 *	5
Y	---	3,6,7, 14,15
X	WO,A,94 16734 (PRESIDENT AND FELLOWS OF HARVARD COLLEGE) 4 August 1994 see the whole document ---	5
Y	J. GEN. VIROL., vol. 73, 1992, pages 3251-3255, XP002016476 ETIENNE-JULIAN ET AL.: "The efficiency of cell targeting by recombinant retroviruses depends on the nature of the receptor and the composition of the artificail cell-virus linker" see the whole document ---	1,3-16
Y	GENE THERAPY, vol. 1, no. supp.2, 1994, page s17 XP002016477 NILSON ET AL.: "Retroviral particles displaying chimaeric MLV envelope proteins" * abstract A64 *	3,7,8
A	---	13-15
Y	WO,A,94 12626 (UNIVERSITY OF MEDICINE AND DENTISTRY OF NEW JERSEY) 9 June 1994 see abstract see page 4 - page 6 see page 9 see claims 1-5 ---	3,4,15
Y	GENE THERAPY, vol. 1, no. supp.2, 1994, page s15 XP002016478 MARIN ET AL.: "Cell targeting by murine recombinant retroviruses" * abstract nr A59 *	1,3-8, 11-16
	---	
	-/--	

# INTERNATIONAL SEARCH REPORT

Int. Patent Application No  
PCI/NL 96/00302

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	PROC. INTERN. SYMP. CONTROL. RELEASE BIOACT. MATER., vol. 20, 25 - 30 July 1993, pages 474-475, XP002016479 TRUONG ET AL.: "A target-specific microspheres drug delivery system made of enzymatically degradable gelatin and chondroitin sulfate coacervates" see the whole document ---	7,12,13
P,X	WO,A,95 26412 (THE UAB RESEARCH FOUNDATION) 5 October 1995 see page 4, line 24 - line 38 see page 5 see page 6, line 1 - line 20 see page 17, line 17 - page 19, line 15 see page 24 - page 25; example 3 ---	3,4,7, 12,15
A	WO,A,93 25234 (THE REGENTS OF THE UNIVERSITY OF CALIFORNIA) 23 December 1993  see the whole document ---	1-4, 6-10, 13-16
A	PROC. NATL ACAD. SCI., vol. 89, no. 13, July 1992, pages 6099-6103, XP002016480 WAGNER ET AL.: "Coupling of adenovirus to transferrin-polylysine/DNA complexes greatly enhances receptor-mediated gene delivery and expression of transfected genes" see the whole document -----	1-16

Form PCT/ISA/210 (continuation of second sheet) (July 1992)

## INTERNATIONAL SEARCH REPORT

national application No.

PCT/NL 96/00302

**Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)**

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:  
Please see Further Information sheet enclosed.
2. ☐ Claims Nos.:  
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

**Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)**

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

## INTERNATIONAL SEARCH REPORT

International Application No. PCT/NL 96/ 00302

FURTHER INFORMATION CONTINUED FROM PCT/ISA/210

Remark : Although claims 1-2 are directed to a method of treatment of (diagnostic method practised on) the human/animal body (as far as they concern an in-vivo method) the search has been carried out and based on the alleged effects of the compound/composition.  
(Art.17.2.a)i) and Rule 39.2.(iv) PCT)

# INTERNATIONAL SEARCH REPORT

Information on patent family members

Int. Application No  
PCT/NL 96/00302

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
FR-A-2649119	04-01-91	NONE	
EP-A-496074	29-07-92	IT-B- 1245748	14-10-94
WO-A-9222332	23-12-92	AT-T- 142506	15-09-96
		AU-A- 2259392	12-01-93
		DE-D- 69213744	17-10-96
		EP-A- 0592564	20-04-94
WO-A-9309221	13-05-93	SE-C- 503225	22-04-96
		CA-A- 2122613	13-05-93
		EP-A- 0614486	14-09-94
		JP-T- 7502026	02-03-95
		SE-A- 9103183	01-05-93
WO-A-9416734	04-08-94	AU-A- 6126794	15-08-94
		EP-A- 0680340	08-11-95
WO-A-9412626	09-06-94	EP-A- 0672129	20-09-95
		JP-T- 8504098	07-05-96
WO-A-9526412	05-10-95	AU-A- 2195495	17-10-95
WO-A-9325234	23-12-93	EP-A- 0650370	03-05-95
		JP-T- 7507689	31-08-95